Notes

Lipo α -Amino- β -hydroxy Acids and **O-Linked Glycosides: Building Blocks for Ceramyl and Glycosphingoyl Peptides**

Michael M. Palian and Robin Polt*

Department of Chemistry, The University of Arizona, Tucson, Årizona 85721

polt@u.arizona.edu

Received June 17, 2001

Introduction

 β -hydroxy α -amino acids, including threonine, serine, and other unusual amino acids, are an important class of chiral, bioactive molecules.¹ In addition to being constituents of bioactive peptides such as cyclosporin,² they are also components of various natural products such as vancomycin and bouvardin. β -Hydroxy α -amino acids are also synthetic intermediates for complex products,³ such as β -lactams,⁴ β -fluoro amino acids,⁵ and aziridines.6

The presence of β -hydroxy amino acids within a peptide allows for additional functionality, e.g., glycosylation.⁷ Glycosides are critical for a number of biological processes, including molecular recognition,8 stability to enzymatic degradation,9 and enhanced transport and biodistribution.¹⁰ In addition, compounds of this type constitute ceramide and glycosylceramide analogues and could provide biologically interesting peptide-glycosphingolipid chimeras.

A number of lipo-amino acid syntheses exist,¹¹ but few possess a side chain capable of glycosylation. The glycosides¹² of differentially protected amino diols¹³ were assembled via three different routes and can be used to create several amphiphilic motifs, including novel gly-

(2) Williams, D. H. Acc. Chem. Res. 1984, 17, 364.

(3) Coppola, G. M., Schuster H. F. Asymmetric Syntheses. Construction of Chiral Molecules Using Amino Acids; John Wiley and Sons: Toronto, 1987.

(4) (a) Miller, M. J. Acc. Chem. Res. 1986, 19, 45-56. (b) Lotz, B.
T.; Miller M. J. J. Org. Chem. 1983, 58, 618-624.
(5) Pansare, S. V.; Vederas, J. C. J. Org. Chem. 1987, 52, 4804-

4810.

(6) Tanner, D. Angew. Chem., Int. Ed. Engl. 1994, 33, 599.

(7) For reviews on glycosylation, see: (a) Whitfield, D. M.; Douglas S. P. *Glycoconjugate J.* **1994**, *13*, 5–17. (b) Toshima, K.; Tatsuta, K. Chem. Rev. 1993. 93. 1503-1531.

(8) Hruby, V. J. Alobeidi, F.; Kazmierski, W.; Biochemistry J. 1990, 268, 249-262.

(9) Rush, B. D.; Ruwart, M. J. J. Med. Chem. 1991, 34, 3140-3143.
(10) Polt, R.; Porreca, R.; Szabò, L. Z.; Bilsky, E. J.; Davis, P.; Davis, T. P.; Horvath, R.; McCormick, J. M.; Yamamura, H. I.; Hruby, V. J.Proc. Natl. Acad. Sci U.S.A. 1994, 91, 7114-7118.

Scheme 1					
tBu-Si-O Me Me N=CPh ₂		1) iBu_5Al_2H $CH_2Cl_2 / -78^\circ$ \longrightarrow tBu 2) RMgBr $-78^\circ \rightarrow RT$		Me J-Si-O Me 2a-2c	OH R N=CPh ₂
	R	Yield	<i>threo</i> Selectivity	Product	
	CH ₃ — CH ₃ (CH ₂) ₅ — CH ₃ (CH ₂) ₉ — CH ₃ (CH ₂) ₁₄ —	60% 60% 72% 62%	> 20:1 > 20:1 > 20:1 >20:1	2a 2b 2c 2d	-

cosphingopeptides. The synthesis represents the first of its type, possessing an O-linked glycoside (as in endogenous glycoproteins) and a C-linked lipid chain to the amino acid, to provide a higher level of enzymatic stability in vivo.

Results and Discussion

In all experiments, D-serine Schiff base 1 was used as the starting material. Reductive alkylation of the methyl ester¹⁴ with ¹Bu₅Al₂H¹⁵ and alkyl Grignard reagents led to protected amino diols **2a**-**d** in enantiomerically pure form and in good yield¹⁶ (Scheme 1). Regardless of chain length, all reductive alkylations proceeded in good yield (60-72%) with excellent diastereoselectivity for the threo product. The products of these reactions were then converted to their respective β -hydroxy amino acids.

In three cases, the reductive alkylation products 2a-cwere hydrogenated and reprotected as the Fmoc carbamates without purification (Scheme 2). This proceeded in 75-82% yield over two steps. The Fmoc-protected silyl amino diols 3a-c were desilylated with BF₃·Et₂O (78-92%) and subjected to selective oxidation of the primary alcohol in the presence of the unprotected secondary alcohol. This was accomplished using the TEMPO oxidation procedure¹⁷ as refined by a group at Merck¹⁸ for substrates containing a single hydroxyl group. We ob-

⁽¹⁾ For recent syntheses of β -hydroxy amino acids, see: (a) Felice, P. D.; Porzi, G.; Sandri, S. Tetrahedron: Asymmetry 1999, 10, 2191-(c) Horikawa, M.; Shigeri, Y.; Yumoto, N.; Yoshikawa, S.;
Nakajimi, T.; Ohfune, Y. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 2027–2032.
(c) Hutton, C. A. Org. Lett. **1999**, *1*, 295–297. (d) Koskinen, A. M. P.;
Hasilla, H.; Myllymaki, V. T.; Rissanen, K. Tetrahedron Lett. **1995**, 36, 5619-5622. (e) Shoa, H.; Goodman, M. J. Org. Chem. 1996, 61, 2582-2583. For reviews, see: (f) Genet, J. P. Pure Appl. Chem. 1996, 68, 593-596. (g) Goleciowski, A.; Jurczak, J. Synlett 1993, 4, 241-5.

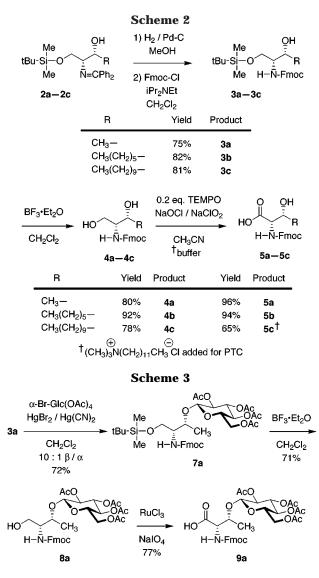
^{(11) (}a) Continantinou-Kototou, V.; Kototos, G. Amino Acids 1999, 16, 273-275. (b) Kototos, G.; Padron, J. M.; Martin, T.; Gibbons, W. A.; Martin, V. S. *J. Org. Chem.* **1998**, *63*, 3741–3744. (c) Kototos, G.; Padron, J. M.; Noula, C.; Gibbons, W. A.; Martin, V. S. *Tetrahedron*: Asymmetry 1996, 7, 857-866. (d) Pignatello, R.; Jansen, G.; Kathmann,

I.; Puglisi, G.; Toth, I. J. Pharm. Sci. 1998, 87, 25–30.
 (12) Peterson, M. A.; Polt, R. J. Org. Chem. 1993, 58, 4309–4314.
 (13) Mitchell, S. A.; Oates, B. D.; Razavi, H. R.; Polt, R. J. Org. Chem. 1998, 63, 8837-8842.

⁽¹⁴⁾ In the hexyl and decyl case, the ethyl ester was also used, and produced no increase in yield, compared to the methyl ester.

⁽¹⁵⁾ Alkylaluminum species aggregate into fluxional trimers in hydrocarbon solution or dimers in more coordinating solvents. NMR studies suggest that the Schiff base esters used in this study react with the ['Bu₃Al]₃ and ['Bu₂AlH]₃ trimers to promote the formation of a dimeric ['Bu₃Al-'Bu₂AlH or 'Bu₅Al₂H] complex prior to reductive alkylation. (a) Eisch, J. J.; Rhee, S. G. J. Organomet. Chem. **1974**, 42, C73. (b) Polt, R.; Peterson, M. A.; DeYoung, L. J. Org. Chem. **1992**, 57, 5469-5480.

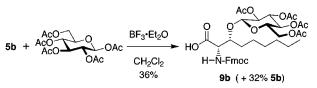
⁽¹⁶⁾ Polt, R.; Peterson, M. A. *Tetrahedron Lett.* **1990**, *31*, 4985–6. (b) Peterson, M. A.; Polt, R. *Synth. Commun.* **1992**, *22*, 477. (c) Sames, D.; Polt, R. Synlett 1995, 552.



served no over-oxidized (diketo) products in the reaction mixtures. With increasing alkyl chain length, it was observed that the oxidation rate slowed, probably due to steric interactions and/or micelle formation. The final step of this sequence proceeded in 65-96% yield. In the decyl case, phase-transfer conditions were required as a result of the high lipophilicity. This provided the protected C₄, C₉, and C₁₃ β -hydroxy lipo amino acids in respectable overall yields, in five steps from the crystalline serine derivative 1.

Glycosylation was achieved directly for protected amino diol **3a** (Scheme 3), using Helferich¹⁹ conditions (HgBr₂), to give the glycoside in 72% yield with greater than 20:1 selectivity for the β -product. Using BF₃·Et₂O, the silyl ether was removed in 71% yield and the primary alcohol oxidized to the acid. However, the TEMPO oxidation was much slower than the simpler cases, proceeding to an unoptimized yield of 43%. Since there is no secondary alcohol in this case, another less selective method of oxidation was employed. Oxidation with RuCl₃/NaIO₄

Scheme 4



provided the Fmoc methyl β -glycosyl amino acid in a 77% yield.

Glycosylation of **3b** and **3c** failed using these same conditions. Similar electrophilic reactions also failed, including the Koenigs-Knorr, Glc(OAc)₅ with BF₃·OEt₂ or TMSOTf, Schmidt's trichloroacetimidate method, as well as tert-butylation, acylation, and benzylation. A priori, these transformations appear to be facile, but from our experience with these compounds the secondary hydroxyl is much more hindered than expected, and its nucleophilicity is reduced due to unfavorable hydrogen bonding with the carbamate NH.²⁰

Therefore, a different route to the other two glycosides was required. The unprotected hexyl β -hydroxy lipo amino acid (**5b**) was directly glycosylated with β -glucose penta-acetate²¹ (Scheme 4). While yields in this reaction were very modest, they were comparable to those seen in similar cases and provided a direct route to the desired glycoside without any further protection or deprotection.

A slightly longer route was devised for synthesis of the $C_{13}\beta$ -hydroxy amino acid glycoside (decyl adduct), which involved a protecting group switch to alleviate some steric hindrance at the secondary hydroxyl (Scheme 5). The reductive alkylation product 2c was subjected to 2 M HCl in THF, resulting in simultaneous cleavage of the Schiff base and the silyl ether to provide the amino diol. This was protected with Fmoc-Cl without purification to yield 4c. This diol was then regioselectively protected at the primary alcohol with benzyl chloroformate to provide 6c. We hoped this compound would be less hindered than silyl-protected 3c and the benzyl carbonate would compete for the carbamoyl N-H hydrogen bond. When the Fmoc alcohol 6c was treated with the trichloroacetimidate and TMSOTf, this glycosylation proceeded in good yield and selectivity for the desired β glycoside **7c**. The benzyl ether was then cleaved by hydrogenolysis and oxidized in two steps via the aldehyde to give the final product (9c) in good yield.

This note describes the stereoselective synthesis of novel lipo-amino acids and three different routes to their glycosides. This approach appears to be the method of choice for the synthesis of these lipid-like glycosyl amino alcohols and acids, and allows for the preparation of lipophilic glycopeptides and peptides, prepared by solidphase methods.²² The resulting amphipathic glycopeptides may possess interesting drug transport activities.²³

^{(17) (}a) Siedlecha, R.; Skazewski, L.; Mlochowski, J. Tetrahedron Lett. 1990, 31, 2177. (b) Rychnovsky, S.; Vaidyananthan, R. J. Org. Chem. 1999, 64, 310-312.

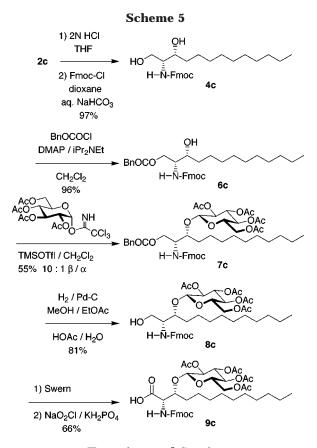
⁽¹⁸⁾ Zhoa, M.; Li, J.; Mano, E.; Song, Z.; Tschaen, D. M.; Grabowski, E. J. J.; Reider, P. J. J. Org. Chem. 1999, 64, 2564–2566. (19) Helferich, B.; Weis, K. Chem Ber. 1956, 89, 314–321.

⁽²⁰⁾ Mitchell, S. A.; Pratt, M. R.; Hruby, V. J.; Polt, R. J. Org. Chem. 2001. 66. 2327-2342

⁽²¹⁾ Salvador, L. A.; Elofsson, M.; Kilhberg, J. Tetrahedron 1995, 51. 5643-5656.

⁽²²⁾ The glycopeptide enkephalin analogue H₂N-Tyr-D-Thr-Gly-Phe-Leu-[9c]-CONH₂ has been synthesized, and its pharmacology is under investigation.

^{(23) (}a) See ref 10. (b) Polt, R.; Mitchell, S. A. In Glycoscience: *Chemistry and Chemical Biology*; Fraser-Reid, B., H. d.; Springer-Verlag: New York, 2000; pp 2355–92. (c) Kriss C. T.; Lou B. S.; Szabò L. Z.; Mitchell, S. A.; Hruby, V. J. Polt, R. Tetrahedron: Asymmetry 2000, 11, 9-25.



Experimental Section

Procedure A. Reductive Alkylation with Grignards (2a–d). Compound 1 was dried overnight in vacuo over P_2O_5 . A solution of 1 (1 equiv in 30–60 mL of CH_2Cl_2) was chilled to –78 °C under argon for 30 min. A solution of Bu_5Al_2H (1.0 equiv, 0.5 M of each in hexanes) was added dropwise via syringe to a stirring solution of 1 over 45 min at –78 °C. Immediately after the $^{3}Bu_5Al_2H$ addition was complete, the alkylmagnesium bromide (3 equiv in Et_2O) was added dropwise via syringe to the stirring solution over 45 min at –78 °C. The solution was allowed to warm to rt and stir overnight. The resulting yellow solution was chilled to 0 °C, carefully quenched with 5 mL of saturated NaHCO₃, and then diluted with 100 mL of CH_2Cl_2 and washed $3 \times$ with saturated NaHCO₃ and $1 \times$ with brine. The organic layer was dried over MgSO₄, filtered, and concentrated in vacuo.

Procedure B. Schiff Base Cleavage and Fmoc Reprotection (3a–c). Compound **2a–c** was dissolved in 100 mL of MeOH before 10% Pd–C (500 mg, regardless of amount of **2**) was added in one portion under Ar. The reaction was vigorously stirred under 1 atm of hydrogen (balloon) at rt. After 2 h, all of the starting material had been consumed, as judged by TLC. The reaction was quenched with 50 mL of CH₂Cl₂, filtered through Celite, and concentrated in vacuo. The crude mixture was then dissolved in dry CH₂Cl₂, and 3 equiv of Pr_2NEt was added in one portion, before Fmoc-Cl (1.0 equiv in 5 mL of CH₂-Cl₂) was added dropwise over 30 min. The reaction was stirred at rt overnight, diluted with 100 mL of CH₂Cl₂, and washed 1× with aqueous HOAc (pH ~3), 3× with saturated NaHCO₃, and 1× with brine. The organic layer was dried over MgSO₄, filtered, and concentrated in vacuo.

Procedure C. Silyl Ether Cleavage (4a–c). Compound **3a–c** was dissolved in 50 mL of dry CH_2Cl_2 . Freshly distilled BF_3 · OEt_2 (6 equiv) was added to the solution in one portion and stirred at rt. After 2 h, all of the starting material had been consumed, as judged by TLC. The reaction was then chilled to 0 °C and quenched with 5 mL of saturated NaHCO₃. The solution was diluted with 100 mL of CH_2Cl_2 and washed 3× with saturated NaHCO₃ and 1× with brine. The organic layer was dried over MgSO₄, filtered, and concentrated in vacuo.

Procedure D. TEMPO Oxidation (5a-c). Compound **4a-c** was dissolved in 3 mL of aqueous buffer (0.67 M KH₂PO₄/Na₂-

HPO₄ buffer) and 3 mL of MeCN and heated to 60 °C in an oil bath. To the rapidly stirring solution was added TEMPO (0.20 equiv) in one portion. Sodium hypochlorite, NaOCl (0.20 equiv, 5.25% commercial bleach dissolved in 0.5 mL of H₂O), and sodium chlorite, NaClO₂ (80%, 2.0 equiv dissolved in 0.5 mL of H₂O), were then added simultaneously over 1 h to the mixture, which was heated at 45 °C for the amount of time specified. The reaction was then acidified to pH 3 with dilute HCl and extracted $3 \times$ with EtOAc. The combined extracts were dried over MgSO₄, filtered, and concentrated in vacuo.

(2*R*,3*R*)-2-(Benzhydrylideneamino)-1-(*tert*-butyldimethylsilanyloxy)butan-3-ol, 2a. Procedure A. The crude reductive alkylation product was chromatographed (4.5% EtOAc/hexanes/0.1% Et₃N, $R_f = 0.40$). Yield: 60% as a yellow oil. IR (cm⁻¹): 3302.2, 3061.4, 2925.5, 2332.7, 1659.6, 1449.7, 1252.1, 1091.6. ¹H NMR (CDCl₃) δ : 7.62 (d, 2H, J = 7.5 Hz), 7.46 (d, 2H, J = 7.5 Hz), 7.23 (m, 6H), 4.01 (m, 1H), 3.70 (ddd, 2H, J = 4.5, 4.5, 10.5, 32.0 Hz), 2.97 (bs, 1H), 2.86 (m, 1H), 1.22 (d, 3H, J = 6.0 Hz), 0.81 (s, 9H), 0.00, 0.00 (s, 6H). ¹³C NMR (CDCl₃) δ : 145.7, 145.4, 128.0, 127.8, 127.3, 127.1, 126.5, 125.9, 75.8, 66.5, 61.2, 25.7, 19.4, 18.1, -5.9. HRMS: calcd for C₂₃H₃₄NO₂Si 384.2359, found 384.2358 (diff = 0.3 ppm).

(2*R*,3*R*)-2-(Benzhydrylideneamino)-1-(*tert*-butyldimethylsilanyloxy)nonan-3-ol, 2b. Procedure A. Crude 2b was chromatographed (2.5% EtOAc/hexanes/0.1% Et₃N, $R_f = 0.40$). Yield: 60% as a yellow oil. IR (cm⁻¹): 3506.0, 3061.4, 2956.4, 2925.5, 2851.46, 2357.46, 1449.7, 1252.14, 1091.6. ¹H NMR δ : 7.70 (d, 2H, J = 7.5 Hz), 7.54 (d, 2H, J = 7.5 Hz), 7.30 (m, 2H), 7.24 (m, 2H), 3.95 (m, 1H), 3.77 (dd, 1H, J = 5.0, 10.5 Hz), 3.72 (dd, 1H, J = 3.0, 10.5 Hz), 3.00 (m, 1H), 1.58 (m, 1 H), 1.47 (m, 3H), 1.33 (m, 4 H), 1.01 (t, 2H, J = 6.5 Hz), 0.92 (t, 3H, J = 7.5 Hz), 7.0, 126.4, 125.9, 79.7, 65.2, 61.7, 43.7, 34.6, 31.6, 29.2, 26.4, 25.8, 22.5, 18.4, 14.0, 0.9, -5.6. HRMS: calcd for C₂₈H₄₄NO₂Si 454.3141, found 454.3154 (diff = 2.7 ppm).

(2*R*,3*R*)-2-(Benzhydrylideneamino)-1-(*tert*-butyldimethylsilanyloxy)tridecan-3-ol. Procedure A. Crude 2c was chromatographed (2.5% EtOAc/hexanes, with 0.1% Et₃N, $R_f = 0.40$). Yield: 72% as a yellow oil. IR (cm⁻¹): 3320.7, 3055.2, 2925.5, 2851.4, 2357.4, 1449.7, 1252.1, 1091.6. ¹H NMR (CDCl₃) δ : 7.69 (d, 2H, J = 7.0 Hz), 7.53 (d, 2H, J = 7.5 Hz), 7.31 (m, 6H), 3.95 (m, 1H), 3.77 (dd, 1H, J = 4.5, 10.3 Hz), 3.71 (dd, 1H, J = 3.0, 10.5 Hz) 2.99 (m, 1H), 1.57, 1.47, 1.31 (m, 18H), 0.93 (t, 3H, J = 6.5 Hz), 0.88 (s, 9H), 0.07, 0.05 (s, 6H). ¹³C NMR (CDCl₃) δ : 145.6, 145.4, 127.9, 127.2, 127.0, 126.4, 125.9, 98.7, 79.7, 65.2, 61.7, 34.6, 31.4, 29.6, 29.5, 29.5, 29.5, 29.3, 26.5, 25.7, 22.6, 14.0, -6.2. HRMS: calcd for C₃₂H₅₂NO₂Si 510.3767, found 510.3678 (diff = 0.2 ppm).

(2*R*,3*R*)-2⁻(Benzhydrylideneamino)-1-(*tert*-butyldimethylsilanyloxy)octadecan-3-ol, 2d. Procedure A. Crude 1d was chromatographed (35% DCM/hex with 0.1% Et₃N, $R_f = 0.24$). Yield: 65.6% as a yellow oil. IR (cm⁻¹): 3320.3, 3055.6, 2924.9, 2852.1, 2357.0, 1449.2, 1252.0, 1089.1. ¹H NMR (CDCl₃): 7.69 (d, J = 7.5 Hz, 2H), 7.54 (d, J = 7.0 Hz, 2H), 7.24 (m, 4H), 3.95 (dt, 1H), 3.73 (ddd, J = 15.0, 5.0, 3.5 Hz, 2H), 2.99 (m, 1H), 1.28 (m, 28H), 0.93 (t, J = 8.0 Hz, 3H), 0.90 (s, 9H), 0.06 (s, 3H), 0.04(s, 3H). ¹³C(CDCl₃):145.8,145.6, 128.0, 127.9, 126.5, 126.0, 79.8, 65.3, 61.8, 34.7, 32.0, 29.7, 29.6, 29.4, 26.6, 25.8, 22.7, 14.1, -5.4. HRMS: (*m*/*z*) obsd = 580.4559, calcd = 580.4557 (diff = 1.9 ppm).

(2*R*,3*R*)-2-(9*H*-Fluoren-9-ylmethoxycarbonylamino)-1-*O* (*tert*-butyldimethylsilyl)butane-1,3-diol, 3a. Procedure B. Crude 3a was chromatographed (20% EtOAc/hexanes $R_f = 0.35$). Yield: 82% as a yellow oil. IR (cm⁻¹): 3438.0, 3061.4, 2925.5, 1702.9, 1511.4, 1252.1, 1103.9. ¹H NMR (CDCl₃) δ : 7.69 (d, 2H, J = 7.5 Hz), 7.53 (t, 2H, J = 7.0 Hz), 7.32 (t, 2H, J = 7.5 Hz) 7.23 (t, 2H, J = 7.5 Hz), 5.38 (d, J = 8.5 Hz), 4.33 (m, 2H), 4.15 (t, 1H, J = 7.0 Hz), 4.12 (m, 1H), 3.78 (ddd, 2H, J = 2.0, 3.5, 10.5, 30.0), 3.48 (m, 1H), 3.20 (bs, 1H), 1.12 (d, 3H, J = 7.0 Hz), 0.83 (s, 9H), 0.00, 0.00 (s, 6H). ¹³C NMR δ : 156.5, 143.9, 141.2, 127.6, 126.9, 125.0, 119.9, 69.0, 66.7, 66.1, 55.2, 47.2, 25.7, 19.8, 17.0, -5.9. HRMS: calcd for C₂₅H₃₆NO₄Si 442.2414, found 442.2435 (diff = 4.9 ppm).

(2*R*,3*R*)-2-(9*H*-Fluoren-9-ylmethoxycarbonylamino)-1-*O*-(*tert*-butyldimethylsilyl)nonane-1,3-diol, 3b. Procedure B. Crude 3b was chromatographed (20% EtOAc/hexanes, $R_f = 0.40$). Yield: 82% as a yellow oil. IR (cm⁻¹): 3431.9, 2950.26,

2925.5, 2857.63, 1696.7, 1503.32, 1449.7, 1252.1, 1103.95. ¹H NMR δ : 7.77 (d, 2H, J = 7.5 Hz), 7.63 (t, 2H, J = 6.5 Hz), 7.40 (t, 2H, J = 7.5 Hz) 7.31 (t, 2H, J = 7.5 Hz), 5.46 (d, J = 8.5 Hz), 4.41 (m, 2H), 4.24 (t, 1H, J = 7.5 Hz), 4.00 (m, 1H), 3.92, 3.85 (dd, 2H, J = 2.0, 10.5 Hz), 3.63 (d, 1H J = 8.0 Hz), 3.28 (bs, 1H), 1.56, 1.45 1.30 (m, 10H), 0.92 (s, 9H), 0.90 (d, 3H, J = 6.5 Hz), 0.00 (s, 6H). ¹³C NMR (CDCl₃) δ : 156.3, 143.9, 141.2, 127.5, 126.9, 125.0, 119.8, 73.1, 66.7, 66.4, 53.8, 47.2, 33.8, 31.6, 29.1, 25.6, 25.4, 18.0, 14.0, -5.2. HRMS: calcd for C₃₀H₄₆NO₄Si 512.3196, found 512.3181 (diff = -2.9 ppm).

(2*R*,3*R*)-2-(9*H*-Fluoren-9-ylmethoxycarbonylamino)-1-*O* (*tert*-butyldimethylsilyl)tridecane-1,3-diol, 3c. Procedure **B.** Crude 3c was chromatographed (17% EtOAc/hexanes, $R_f =$ 0.40). Yield: 81% as a yellow oil. IR (cm⁻¹): 3438.0, 3073.7, 2925.5, 2851.4, 1696.7, 1505.3, 1449.7, 1252.1, 1103.9. ¹H NMR (CDCl₃) &: 7.77 (d, 2H, J = 7.5 Hz), 7.61 (t, 2H, J = 7.5 Hz), 7.40 (t, 2H, J = 7.5 Hz), 7.31 (t, 2H, J = 7.5 Hz), 5.46 (d, J = 9.0Hz), 4.39 (m, 2H), 4.24 (t, 1H, J = 7.5 Hz), 3.99 (t, 1H, J = 5.0Hz), 3.92 (dd, 1H, J = 3.0, 10.5 Hz), 3.84 (dd, 1H, J = 2.0, 10.5 Hz), 1.55 (m, 2H), 1.28–1.20 (m, 16H), 0.91 (s, 9H), 0.87 (t, 3H, J = 7.5 Hz), 0.08 (s, 3H), 0.07 (s, 3H). ¹³C NMR (CDCl₃) &: 156.3, 143.9, 141.2, 127.5, 126.9, 125.0, 119.9, 73.2, 66.7, 66.5, 53.8, 47.3, 33.8, 31.8, 29.5, 29.5, 29.2, 25.7, 25.5, 22.6, 18.0, 14.0, -5.6. HRMS: calcd for C₃₄H₅₄NO₄Si 568.3822, found 568.3839 (diff = 2.9 ppm).

(2*R*, 3*R*)-2-(9*H*-Fluoren-9-ylmethoxycarbonylamino)butane-1,3-diol, 4a. Procedure C. Crude 4a was chromatographed (50% EtOAc/hexanes, $R_f = 0.35$). Yield: 80% as a white solid. ¹H NMR (CDCl₃/MeOH) δ : 7.75 (d, 2H, J = 8.0 Hz), 7.62 (d, 2H, J = 7.5 Hz), 7.39 (t, 2H, J = 7.5 Hz), 7.31 (t, 2H, J = 8.0 Hz), 5.96 (d, 1H, J = 9.0 Hz), 4.44 (dd, 1H, J = 7.0 Hz), 4.37 (dd, 1H, J = 7.0, 10.5 Hz), 4.21 (t, 1H, J = 7.0 Hz), 4.06 (m, 1H), 3.75 (bs, 1H), 3.65 (d, 2H, J = 5.5 Hz), 3.52 (m, 1H), 1.17 (d, 3H, J = 6.5 Hz). ¹³C NMR (CDCl₃/CD₃OD) δ : 157.2, 143.5, 141.0, 127.4, 126.7, 124.7, 119.6, 66.4, 66.3, 62.5, 56.6, 48.4, 46.9, 19.3. HRMS: calcd for C₁₉H₂₂NO₄ 328.1549, found 328.1552 (diff = 1.0 ppm).

(2*R*, 3*R*)-2-(9*H*-Fluoren-9-ylmethoxycarbonylamino)nonane-1,3-diol, 4b. Procedure C. Crude 4b was chromatographed (50% EtOAc/hexanes, $R_f = 0.35$). Yield: 92% as a light yellow oil. IR (cm⁻¹): 3438.0, 2931.7, 1641.17, 1233.1, 1036.0. ¹H NMR (CDCl₃) δ : 7.77 (d, 2H, J = 7.5 Hz), 7.60 (d, 2H, J =7.5 Hz), 7.40 (t, 2H, J = 7.5 Hz), 7.31 (t, 2H, J = 7.5 Hz), 5.48 (d, 1H, J = 8.0 Hz), 4.43 (m, 2H), 4.22 (t, 1H, J = 7.0 Hz), 3.93 (t, 1H, J = 6.5 Hz), 3.81 (m, 2H), 3.64 (t, 1H, J = 6.5 Hz), 1.48 (m, 2H), 1.28–1.20 (m, 8H), 0.92 (dd, 1H, J = 7.0, 9.5 Hz), 0.87 (t, 3H, J = 7.0 Hz). ¹³C NMR (CDCl₃) δ : 156.2, 143.8, 141.3, 127.6, 127.0, 124.9, 119.9, 72.5, 66.7, 64.9, 54.8, 47.2, 34.1, 31.7, 29.1, 25.5, 22.5, 22.1, 13.9. HRMS: calcd for C₂₄H₃₂NO₄ 398.2331, found 398.2325 (diff = 1.6 ppm).

(2*R*,3*R*)-2-(9*H*-Fluoren-9-ylmethoxycarbonylamino)tridecane-1,3-diol, 4c. Procedure C. Crude 4c was chromatographed (50% EtOAc/hexanes, $R_f = 0.40$). Yield: 78% as a white solid. IR (cm⁻¹): 3345.4, 3061.4, 2919.3, 2845.2, 1690.5, 1536.1, 1449.7, 1252.1, 1079.2. ¹H NMR (CDCl₃) δ : 7.76 (d, 2H, J = 8.0Hz), 7.60 (d, 2H, J = 7.0 Hz), 7.40 (t, 2H, J = 7.5 Hz), 7.31 (t, 2H, J = 7.5 Hz), 5.50 (d, 1H, J = 8.0 Hz), 4.43 (dd, 2H, J = 7.0, 15.0 Hz), 4.22 (t, 1H, J = 7.0 Hz), 3.92 (t, 1H, J = 6.5 Hz), 3.81 (bs, 2H), 3.64 (bs, 1H), 1.48 (m, 2H), 1.28–1.20 (m, 16H), 0.88 (t, 3H, J = 7.0 Hz). ¹³C NMR (CDCl₃) δ : 156.8, 143.8, 141.3, 127.6, 127.0, 125.0, 119.9, 72.8, 66.7, 65.2, 54.6, 47.2, 34.1, 31.8, 29.5, 29.5, 29.5, 29.5, 29.3, 25.5, 22.6, 14.0. HRMS: calcd for C₂₈H₄₀NO₄ 454.2957, found 454.2957 (diff = 0.1 ppm).

Alternate Procedure for (2R,3R)-2-(9H-Fluoren-9-ylmethoxycarbonylamino)tridecane-1,3-diol, 4c. Compound 2c (161 mg, 0.3163 mmol) was dissolved in 2 M HCl (3 mL) and THF (3 mL) and stirred at rt for 2 h. Upon completion, as judged by TLC, the crude reaction mixture was made basic (pH \approx 8.5) with solid NaHCO₃. Fmoc-Cl (108 mg, 0.4174 mmol, 1.3 equiv in 1 mL dioxane) was then added via syringe to the stirring mixture over 30 min at rt and allowed to react overnight. The mixture was diluted with 50 mL of EtOAc and washed 3× with saturated NaHCO₃ and 1× with brine. The organic layer was dried over MgSO₄, filtered, and concentrated in vacuo. Crude 4c was chromatographed (50% EtOAc/hexanes, $R_f = 0.40$). Yield: 110.1 mg (77% over three transformations, as a light yellow oil). (2.5,3*R*)-(9*H*-Fluoren-9-ylmethoxycarbonylamino)-3-hydroxybutanoic Acid, 5a. Procedure D. Total reaction time was 12 h. Crude 5a was chromatographed (gradient: 100% EtOAc, followed by EtOAc with 0.5% HOAc, $R_f = 0.60$). Yield: 96% as a white solid. IR (cm⁻¹): 3376.3, 2944.0, 1752.3, 1517.6, 1369.4, 1221.2, 1042.2. ¹H NMR δ : 7.77 (d, 2H, J = 7.0 Hz), 7.61 (dd, 2H, J = 7.5, 11.5 Hz), 7.33 (t, 2H, J = 7.5), 7.26 (t, 2H, J = 7.5 Hz), 4.32 (1H, d, J = 7.5 Hz), 4.30 (2H, m), 4.17 (1H, m), 4.15 (1H, t, J = 7.5 Hz), 1.19 (3H, d, J = 6.5 Hz). ¹³C NMR (MeOH) δ : 174.5, 158.8, 145.0, 142.4, 128.7, 128.0, 126.1, 120.8, 68.5, 68.0, 61.0, 48.2, 20.3. HRMS: calcd for C₁₉H₂₀NO₅ 342.1341, found 342.1352 (diff = 3.1 ppm).

(2.S,3*R*)-2-(9*H*-Fluoren-9-ylmethoxycarbonylamino)-3hydroxynonanoic acid, 5b. Procedure D. Total reaction time 24 h. Crude 5b was chromatographed (gradient: 100% EtOAc, followed by EtOAc with 0.5% HOAc, $R_f = 0.65$). Yield: 94% as a yellow oil. IR (cm⁻¹): 3444.2, 2931.7, 2061.0, 1690.5, 1641.1, 1517.6, 1227.4, 1054.5. ¹H NMR (CDCl₃) δ : 7.73 (d, 2H, J = 7.5Hz), 7.58 (t, 2H, J = 8.5 Hz), 7.37 (m, 2H), 7.27 (t, 2H, J = 7.5Hz), 5.95 (1H, d, J = 8.5 Hz), 4.42 (1H, d, J = 8.5 Hz), 4.36 (2H, d, J = 7.5 Hz), 4.23 (1H, t, J = 6.5 Hz), 4.19 (1H, t, J = 7.5 Hz), 1.52 (2H, m), 1.26 (10H, m), 0.85 (3H, t, J = 7.5 Hz). ¹³C NMR (CDCl₃) δ : 175.0, 157.8, 143.6, 141.2, 128.2, 127.5, 125.0, 120.2, 71.7, 67.4, 57.8, 46.9, 33.2, 31.5, 29.0, 25.4, 22.3, 13.9. HRMS: calcd for C₂₄H₃₀NO₅ 412.2124, found 412.2127 (diff = 0.8 ppm).

(2.5,3*R*)-2-(9*H*-Fluoren-9-ylmethoxycarbonylamino)-3hydroxytridecanoic Acid, 5c. Procedure D. Total reaction time was 24 h. Crude 5c was chromatographed (gradient: 100% EtOAc, followed by EtOAc with 0.5% HOAc, $R_f = 0.70$). Yield: 66% as a yellow oil. IR (cm⁻¹): 3357.8, 3607.5, 2925.5, 2851.4, 1721.4, 1530.0, 1449.7, 1252.1, 1085.4. ¹H NMR (CDCl₃/MeOH) δ : 7.75 (d, 2H, J = 8.0 Hz), 7.63 (t, 2H, J = 8.5 Hz), 7.36 (t, 2H, J = 7.5 Hz), 7.28 (t, 2H, J = 7.5 Hz), 6.53 (d, 1H, J = 9.5 Hz), 4.39 (dd, 1H, J = 7.5, 11.0 Hz), 4.34 (dd, 1H, J = 7.0, 11.0 Hz), 4.27 (bs, 1H), 4.22 (t, 1H, J = 7.0 Hz), 4.11 (t, 1H, J = 6.5), 1.49 (t 2H, J = 8.0), 1.28–1.20 (m 16H), 0.85 (t, 3H, J = 7.5 Hz). ¹³C NMR (CDCl₃/MeOH) δ : 173.5, 157.3, 143.2, 140.5, 127.0, 126.2, 124.2, 119.2, 70.6, 66.6, 57.7, 47.2, 33.2, 31.1, 28.7, 28.7, 28.7, 28.7, 24.8, 21.8, 12.8. HRMS: calcd for C₂₈H₃₈NO₅ 468.2750, found 468.2745 (error = -1.0 ppm).

(2R,3R)-Carbonic Acid Benzyl Ester 2-(9H-Fluoren-9ylmethoxycarbonylamino)-3-hydroxytridecyl Ester, 6c. Diol 4c (930.0 mg, 2.052 mmol) and DMAP (36.1 mg, 14.4 mol %) were placed in a flame-dried flask. CH₂Cl₂ (25 mL) was added, and the resulting suspension was chilled to -78 °C. iPr_2NEt (700 μ L, 1.96 equiv) and benzylchloroformate (300 μ L, 1.02 equiv) were then added sequentially to the stirring suspension. The mixture was slowly allowed to warm to rt and reacted for a total of 48 h. The product was diluted with 50 mL of CH₂Cl₂ and washed $1 \times$ with dilute HCl (pH \sim 3), $3 \times$ with saturated NaHCO₃, and $1 \times$ with brine. The organic layer was dried over MgSO₄, filtered, and concentrated in vacuo. Crude 4c was chromatographed (30% EtOAc/hexanes, $R_f = 0.45$). Yield: 1.09 g, 96% as a white solid. IR (cm⁻¹): 3376.3, 2944.0, 1752.3, 1517.6, 1369.4, 1221.2, 1042.2. ¹H NMR (CDCl₃) δ : 7.75 (d, 2H, J = 7.5Hz), 7.57 (d, 2H, J = 7.5, 11.5 Hz), 7.35 (m, 6H), 7.29 (t, 2H, J = 7.5 Hz), 5.19 (d, 1H, J = 9.0 Hz), 5.15 (s, 2H), 4.40 (2H, d, J = 6.0 Hz), 4.29 (1H, dd, J = 11.5, 7.0 Hz), 4.21 (1H, dd, J =11.5, 7.0 Hz), 4.20 (1H, t, J = 7.5 Hz), 3.87 (1H, d, J = 6.5 Hz), 3.74 (1H, bs), 1.42 (4H, m), 1.24 (14H, m), 0.87 (3H, t, J = 6.5Hz). ¹³C NMR (CDCl₃) δ: 156.4, 155.3, 143.8, 141.3, 134.9, 128.6, 128.3, 127.6, 127.0, 125.0, 119.9, 70.0, 69.8, 67.1, 66.8, 53.1, 47.2, 33.7, 31.8, 29.5, 29.5, 29.4, 29.3, 25.6, 22.6, 14.0.

(2*R*,3*R*)-2-(9*H*-Fluoren-9-ylmethoxycarbonylamino)-1-*O* (*tert*-butyldimethylsilyl)-3-*O*-(2,3,4,6-tetra-*O*-acetylglucosyl)butane-1,3-diol, 7a. In a flame-dried flask, the Fmoc acceptor 3a (80.1 mg, 0.1813 mmol), acetobromoglucose (120.0 mg, 0.2919 mmol, 1.61 equiv), and powdered 3 Å molecular sieves (200 mg) were dissolved in CH_2Cl_2 and the mixture chilled to 0 °C. A mixture of $Hg(CN)_2$ (60.0 mg, 0.2375 mmol, 1.30 equiv) and $HgBr_2$ (81.1 mg, 0.2250 mmol, 1.24 equiv) was added portionwise over 45 min by solid-addition funnel to the stirring suspension. The suspension was then allowed to warm to rt and stirred for a total of 40 h. After being quenched with Et_3N (0.2 mL), the solution was diluted with 50 mL of CH_2Cl_2 and filtered through Celite. The mixture was then washed 1× with saturated aqueous Na_2SO_4 , 1× with saturated aqueous $NaHCO_3$, and 1× with brine, dried with MgSO₄, filtered, and concentrated in vacuo. Crude **7a** was chromatographed (35% EtOAc/hexanes, $R_f = 0.40$). Yield: 101 mg, 72%, as a yellow oil. IR (cm⁻¹): 3067.5, 2950.2, 1752.3, 1511.4, 1369.4, 1221.2, 1042.2. ¹H NMR (CDCl₃) δ : 7.73 (d, 2H, J = 8.0 Hz), 7.60 (dd, 2H, J = 7.5, 4.5 Hz), 7.37 (t, 2H, J = 7.5 Hz), 7.29 (m, 2H), 5.21 (t, 1H, J = 9.5 Hz), 5.06 (t, 1H, J = 9.5 Hz), 5.03 (d, 2H, J = 8.0 Hz), 4.94 (t, 1H, J = 8.5 Hz), 4.52 (d, 1H, J = 8.0 Hz), 4.35 (t, 1H, J = 6.5 Hz), 4.20 (m, 1H), 4.13 (dd, 1H, J = 2.0, 6.0 Hz), 4.07 (dd, 1H, J = 2.0, 12.5 Hz), 3.64 (m, 2H), 3.63 (t, 1H, J = 8.0 Hz), 3.57 (dd, 1H, J = 4.0, 9.0 Hz), 2.02, 2.00, 2.00, 1.99 (s, 12H), 1.14 (d, 3H, J = 6.5 Hz), 0.85 (s, 9H), 0.03, 0.02 (s, 6H). ¹³C NMR (CDCl₃) δ : 170.1, 170.0, 169.4, 169.3, 156.4, 143.9, 141.2, 127.6, 127.0, 125.1, 119.9, 98.3, 72.4, 72.0, 71.8, 71.4, 68.3, 66.6, 61.8, 56.2, 47.2, 25.7, 20.6, 20.6, 20.5, 20.5, 18.1, 16.3, -5.9. HRMS: calcd for C₃₉H₅₄NO₁₃-Si 772.3364, found 772.3370 (error = 0.7 ppm).

(2R,3R)-Carbonic Acid Benzyl Ester 2-(9H-Fluoren-9ylmethoxycarbonylamino)-3-O-(2,3,4,6-tetra-O-acetylglucosyl)tridecyl Ester, 7c. To a flame-dried flask were added acceptor 6c (31.3 mg, 0.0576 mmol) and tetra-O-acetyl-glucose $\alpha\text{-trichloroacetimidate}$ (69.1 mg, 2.44 equiv), and the mixture was azeotroped $2 \times$ with PhCH₃. Molecular sieves (4 Å, ~100 mg) were added to the mixture before it was dissolved in CH2- Cl_2 (6 mL) and chilled to 0 °C. TMSOTf (23 μ L, 2.20 equiv) was then added to the stirring solution dropwise over 10 min. The mixture was stirred and allowed to warm to rt overnight. The reaction was filtered through Celite, washed three times with saturated aqueous NaHCO₃, dried with MgSO₄, filtered again, and concentrated in vacuo. The crude was chromatographed (25% EtOAc/hexanes, $R_f = 0.35$). Yield: 27.7 mg (55%, as brittle white foam). ¹H NMR (CDCl₃/CD₃OD) δ : 7.77 (d, 2H, J = 7.5Hz), 7.62 (t, 2H, J = 6.0 Hz), 7.40 (t, 2H, J = 7.5 Hz), 7.33 (m, 8H), 5.97 (d, 1H, J = 9.0 Hz), 5.23 (t, 1H, J = 9.5 Hz), 5.13 (d, 2H, J = 8.0 Hz), 5.06 (t, 1H, J = 10.0 Hz), 4.97 (t, 1H, J = 9.5Hz), 4.56 (d, 1H, J = 8.0 Hz), 4.37 (t, 2H, J = 6.0 Hz), 4.29 (dd, 1H, J = 7.0, 10.0 Hz), 4.22 (t, 1H, J = 7.0 Hz), 4.16 (dd, 1H, J= 7.0, 12.0 Hz), 4.16 (m, 1H), 4.10 (dd, 1H, J = 2.5, 12.0 Hz), 4.05 (m, 1H,), 3.78 (m, 1H,), 3.72 (m, 1H), 2.08, 2.05, 2.04, 2.03 (s, 12H), 1.57 (m, 2H), 1.25 (m, 16H, J = 6.4 Hz), 0.87 (t, 3H, J= 7.5 Hz), ¹³C NMR (CDCl₃/CD₃OD) δ : 171.5, 170.8, 170.2, 170.1, 157.2, 144.1, 141.5, 128.8, 127.9, 125.3, 120.1, 100.5, 78.8, 73.2, 73.0, 72.0, 71.8, 70.2, 70.0, 68.7, 62.1, 47.4, 32.1, 32.0, 29.5, 29.3, 29.1, 25.8, 23.5, 20.4, 20.4, 14.9.

(2R,3R)-2-(9H-Fluoren-9-ylmethoxycarbonylamino)-3-O-(2,3,4,6-tetra-O-acetylglucosyl)butane-1,3-diol, 8a. To a stirring solution of (1.56 g, 2.021 mmol, in 20 mL of CH₂Cl₂) was added by syringe freshly distilled BF3. OEt2 (1.53 mL, 12.129 mmol, 6.0 equiv) in one portion and the mixture stirred at rt for 2 h. The reaction was then chilled to 0 °C before 3 mL of saturated aqueous NaHCO₃ was added to quench the reaction. The mixture was then washed three times with saturated aqueous NaHCO₃, dried with MgSO₄, filtered, and concentrated in vacuo. The crude was chromatographed (66% EtOAc/hexanes, $R_f = 0.35$). Yield: 870 mg (66%, as brittle white foam). IR (cm⁻¹): 3342.2, 2950.2, 1752.3, 1511.4, 1375.6, 1221.2, 1036.0. ¹H NMR (CDCl₃) δ : 7.71 (d, 2H, J = 7.5 Hz), 7.56 (d, 2H, J = 7.0 Hz), 7.35 (t, 2H, J = 7.5 Hz), 7.27 (m, 2H), 5.19 (t, 1H, J = 9.5 Hz), 5.08 (d, 2, J = 9.0 Hz), 4.99 (t, 1H, J = 9.5 Hz), 4.91 (t, 1H, J = 0.5 Hz), 4.91 (t, 1H, J = 0.0 (t, 1H 9.0 Hz), 4.48 (d, 1H, J = 8.0 Hz), 4.38 (t, 1H, J = 8.5 Hz), 4.32 (t 1H, J = 7.0 Hz), 4.22 (d, 1H, J = 11.5 Hz), 4.18 (t, 1H, J = 7.0Hz), 4.07 (m, 1H), 3.67 (m, 2H), 3.59 (t, 1H, J = 5.7 Hz), 2.03, 1.99, 1.99, 1.97 (s, 12H), 1.12 (d, J = 6.0 Hz). ¹³C NMR (CDCl₃) $\delta:\ 170.6,\ 170.1,\ 169.3,\ 169.3,\ 156.6,\ 143.8,\ 141.2,\ 127.6,\ 127.0,$ 125.1, 119.9, 99.3, 77.3, 74.2, 72.4, 71.9, 71.1, 68.5, 66.7, 62.1, 61.8, 55.7, 47.2, 20.5, 20.5, 20.5, 20.5, 16.7. HRMS: calcd for $C_{33}H_{40}NO_{13}$ 658.2500, found 658.2502 (error = 0.4 ppm).

(2*R*,3*R*)-2-(9*H*-Fluoren-9-ylmethoxycarbonylamino)-3-*O* (2,3,4,6-tetra-*O*-acetylglucosyl)tridecane-1,3-diol, 8c. Benzyl carbonate 7c (22.4 mg, 0.0256 mmol) was placed in a roundbottom flask. To this were added MeOH (5 mL), EtOAc (5 mL), aqueous HOAc (0.5 mL, pH ~3.5), and Pt-C (24.0 mg). The reaction was purged with H₂ four times and stirred under 1 atm of H₂ until judged complete by TLC (2 h). The reaction was quenched with 20 mL of CH₂Cl₂, filtered through Celite, and washed three times with saturated aqueous NaHCO₃, dried with MgSO₄, filtered again, and concentrated in vacuo. The crude was chromatographed (54% EtOAc/hexanes, $R_f = 0.45$). Yield: 16.2 mg (80.6%, as a colorless oil). ¹H NMR (CDCl₃) δ : 7.76 (d, 2H, J = 7.5 Hz), 7.60 (m, 2H,), 7.38 (t, 2H, J = 8.0 Hz), 7.32 (q, 2H, J = 7.5 Hz), 5.23 (t, 1H, J = 9.5 Hz), 5.02 (t, 1H, J = 9.0 Hz), 4.99 (t, 1H, J = 9.0 Hz), 4.56 (d, 1H, J = 8.0 Hz), 4.38 (t, 2H, J = 7.0 Hz), 4.29 (dd, 1H, J = 7.0, 10.0 Hz), 4.23 (t, 1H, J = 7.0 Hz), 4.16 (dd, 1H, J = 7.0, 12.0 Hz), 4.05 (m, 1H,), 3.78 (d, 1H, J = 8.0 Hz), 3.73 (m, 1H, J = 9.5 Hz), 2.09, 2.05, 2.05, 2.02 (s, 21H), 1.57 (m, 2H), 1.25 (m, 16H, J = 6.5 Hz), 0.87 (t, 3H, J = 7.0 Hz). HRMS: calcd for C₄₂H₅₈NO₁₃ 784.3908, found 784.3912 (error = 0.5 ppm).

(2*S*,3*R*)-2-(9*H*-Fluoren-9-ylmethoxycarbonylamino)-3-*O*-(2,3,4,6-tetra-O-acetylglucosyl)butanoic Acid, 9a. Protected glycoside 7a (61.6 mg, 0.0936 mmol) was dissolved in aqueous MeCN (6 mL, 50%) before NaIO₄ (200 mg, 0.9350 mmol, 9.98 equiv) and RuCl₃·H₂O (3.0 mg, 0.0144 mmol, 15 mol %) were added in one portion. The resulting mixture was stirred at rt for 4 h. Once all of the starting material had been consumed, the reaction was quenched with PrOH (20 mL) and stirred for an additional 2 h. The mixture was then filtered through Celite, washed twice with saturated aqueous sodium sulfate, washed one time with brine, dried with MgSO₄, filtered, and concentrated in vacuo. The crude was chromatographed (gradient: 100% EtOAc, followed by 100% EtOAc with 0.5% HOAc, $R_f =$ 0.65). Yield: 48.9 mg (77%, as a white solid). IR (cm⁻¹): 3376.3, 2944.0, 1752.3, 1517.6, 1369.4, 1221.27, 1042.2. ¹H NMR δ: 7.76 (d, 2H, J = 8.0 Hz), 7.63 (t, 2H, J = 8.0 Hz), 7.39 (t, 2H, J = 7.5Hz), 7.32 (m, 2H), 5.66 (d, 1H, J = 9.0 Hz), 5.19 (t, 1H, J = 9.5 Hz), 5.09 (t, 1H, J = 9.5 Hz), 4.95 (t, 1H, J = 8.0 Hz), 4.54 (d, 1H, J = 8.0 Hz), 4.43 (d, 1H, J = 8.0 Hz), 4.39 (m, 2H), 4.39 (m, 1H), 4.38 (dd, 1H, J = 2.0, 6.0 Hz), 4.25 (t, 1H, J = 7.5 Hz), 4.09 (dd, 1H, J = 3.5, 12.5 Hz), 3.65 (d, 1H, J = 9.5 Hz), 2.10, 2.04, 2.03, 2.01 (s, 12H), 1.23 (d, 3H, J = 6.5 Hz). ¹³C NMR δ : 172.4, 171.7, 170.2, 169.3, 169.2, 156.7, 143.7, 141.1, 127.6, 127.0, 125.1, 119.8, 99.4, 75.8, 72.5, 71.5, 71.0, 68.3, 67.2, 61.5, 58.0, 47.0, 29.5, 20.7, 20.5, 20.5, 20.3, 17.5. HRMS: calcd for C33H38NO14 672.2292, found 672.2288 (error = -0.7 ppm).

(2S,3R)-2-(9H-Fluoren-9-ylmethoxycarbonylamino)-3-O-(2,3,4,6-tetra-O-acetylglucosyl)nonanoic acid, 9b. Threonine analogue **5b** (63.3 mg, 0.1540 mmol) and β -glucose penta-acetate (71.0 mg, 0.1820 mmol, 1.18 equiv) were placed in a 50 mL round-bottom flask and azeotroped $2 \times$ with toluene. The mixture was dissolved in CH_2Cl_2 and treated with $BF_3 \cdot OEt_2$ (60 μL , 0.4739 mmol, 3.07 equiv). The solution was then stirred at rt for 36 h. After being judged complete by TLC, the reaction mixture was diluted with 15 mL of CH₂Cl₂ and quenched with 1 mL of sat. NaHCO₃. The mixture was then washed $1 \times$ with brine, dried with MgSO₄, filtered, and concentrated in vacuo. The crude was chromatographed (5% MeOH/95% CH₂Cl₂ with 0.4% HOAc, $R_f = 0.35$). Yield: 40.2 mg (36%, as a yellow oil). ¹H NMR (CDCl₃/CD₃OD/toluene- d_8) δ : 7.75 (d, 2H, J = 7.5 Hz), 7.62 (t, 2H, J = 8.0 Hz), 7.39 (t, 2H, J = 7.5 Hz), 7.31 (t, 2H, J= 7.5 Hz), 5.60 (d, 1H, J = 9.0 Hz), 5.18 (t, 1H, J = 9.5 Hz) 5.10 (t, 1H, J = 10.0 Hz), 4.99 (t, 1H, J = 9.0 Hz), 4.56 (d, 1H, J =8.0 Hz), 4.47 (m, 1H,), 4.38 (m, 2H), 4.25 (m, 1H), 4.20 (m, 1H), 4.09 (m, 2H), 3.62 (d, 1H, J = 10.5 Hz), 2.10, 2.03, 2.02, 2.00 (s, 12H), 1.58, 1.50 (m, 2H), 1.25 (m, 8H), 0.87 (t, 3H, J = 6.5 Hz). ¹³C NMR (CDCl₃/MeOH/PhCH₃) δ: 172.0, 171.8, 170.2, 169.3, 169.1, 156.7, 143.3, 141.2, 128.8, 128.1, 125.2, 119.9, 100.9, 80.7, 72.7, 71.7, 71.2, 68.3, 67.6, 67.4, 61.4, 56.8, 47.2, 29.6, 25.3, 22.5, 21.4, 20.9, 13.9. HRMS: calcd for C₃₈H₄₇NO₁₄Na 764.2889, found 764.2909 (error = 2.6 ppm).

(2S,3R)-2-(9H-Fluoren-9-ylmethoxycarbonylamino)-3-O-(2,3,4,6-tetra-O-acetylglucosyl)tridecanoic Acid, 9c. Glycoside 8a (17.2 mg, 0.02193 mmol) was azeotroped with toluene $\times 2$. One milliliter of CH₂Cl₂ was added to a round-bottom flask and chilled to -40 °C. Oxayl chloride (15 μ L, 0.1713 mmol, 7.8 equiv) and DMSO (15 $\mu L,~0.2111$ mmol, 9.6 equiv) were sequentially added in an inert atmosphere, and the mixture was stirred for 2 min. Compound 8a was then dissolved in 1 mL of CH_2Cl_2 and added via syringe. The mixture was stirred at -40°C for 30 min prior to the addition of Et₃N (51 μ L, 0.3635 mmol, 16.5 equiv). The reaction was allowed to warm to rt and stirred for 1 h until judged complete by TLC. The completed reaction was then diluted with 10 mL of CH_2Cl_2 , washed with saturated aqueous $NH_4Cl \times 4$, and concentrated in vacuo. The residue was redissolved in 'BuOH (5 mL) and EtOAc (5 mL), charged with NaClO₂ (46 mg, 0.3376 mmol, 15.0 equiv) and KH₂PO₄ (55.2 mg,

0.4029 mmol, 18.3 equiv), and stirred for 30 min at rt. The crude was chromatographed (5% MeOH/ 95% CH₂Cl₂ with 0.2% HOAc, R_{f} = 0.35). Yield: 11.3 mg (65%, as a yellow oil). ¹H NMR (CDCl₃/MeOH) δ : 7.74 (d, 2H, J = 8.0 Hz), 7.62 (t, 2H, J = 6.5 Hz), 7.37 (t, 2H, J = 7.5 Hz), 7.29 (t, 2H, J = 7.5 Hz), 5.88 (d, 1H, J = 9.5 Hz), 5.16 (t, 1H, J = 9.5 Hz), 5.09 (t, 1H, J = 10.0 Hz), 4.97 (t, 1H, J = 8.5 Hz), 4.56 (d, 1H, J = 8.0 Hz), 4.37 (t, 1H, J = 7.5 Hz), 3.24 (d, 1H, J = 9.5 Hz), 2.05, 2.03, 2.00, 1.99 (s, 12H), 1.58 (m, 2H), 1.25 (m, 16H, J = 6.5 Hz), 0.85 (t, 3H, J = 8.0 Hz), HzMS: calcd for C₄₂H₅₅NO₁₄Na 820.3515, found 820.3478 (error = 4.5 ppm).

Acknowledgment. We thank the A.R.C.S. Foundation (Phoenix Chapter – Spetzler Fellowship), the U.S. Army (DAMD17-99-1-9539), the National Science Foundation (CHE-9526909 and CHE-920112), and NIDA (DA-06284) for partial funding.

Supporting Information Available: NMR spectra for obtained compunds. This material is available free of charge via the Internet at http://pubs.acs.org.

JO015844V